

Pharmacological effects of biotin[☆]

Cristina Fernandez-Mejia*

Nutritional Genetics Unit, Biomedical Research Institute, National University of Mexico/Pediatric National Institute, Mexico City, CP 04530, Mexico

Received 30 March 2005; received in revised form 30 March 2005; accepted 30 March 2005

Abstract

In the last few decades, more vitamin-mediated effects have been discovered at the level of gene expression. Increasing knowledge on the molecular mechanisms of these vitamins has opened new perspectives that form a connection between nutritional signals and the development of new therapeutic agents. Besides its role as a carboxylase prosthetic group, biotin regulates gene expression and has a wide repertoire of effects on systemic processes. The vitamin regulates genes that are critical in the regulation of intermediary metabolism: Biotin has stimulatory effects on genes whose action favors hypoglycemia (insulin, insulin receptor, pancreatic and hepatic glucokinase); on the contrary, biotin decreases the expression of hepatic phosphoenolpyruvate carboxykinase, a key gluconeogenic enzyme that stimulates glucose production by the liver. The findings that biotin regulates the expression of genes that are critical in the regulation of intermediary metabolism are in agreement with several observations that indicate that biotin supply is involved in glucose and lipid homeostasis. Biotin deficiency has been linked to impaired glucose tolerance and decreased utilization of glucose. On the other hand, the diabetic state appears to be ameliorated by pharmacological doses of biotin. Likewise, pharmacological doses of biotin appear to decrease plasma lipid concentrations and modify lipid metabolism. The effects of biotin on carbohydrate metabolism and the lack of toxic effects of the vitamin at pharmacological doses suggest that biotin could be used in the development of new therapeutics in the treatment of hyperglycemia and hyperlipidemia, an area that we are actively investigating.

© 2005 Elsevier Inc. All rights reserved.

Keywords: Biotin; Carbohydrates; Lipids; Gene expression; Carboxylases; Glucokinase

In the last few decades, more vitamin-mediated effects have been discovered at the level of gene expression [1–3] in addition to their well-known roles as substrates and cofactors [4]. The best recognized examples are the lipophilic Vitamins A and D that serve as ligand precursors of the hormone nuclear receptors superfamily and thus affect systemic functions such as morphogenesis, immunity, growth and epithelial cell differentiation [1–3]. Increasing knowledge on the molecular mechanisms of these vitamins has opened new perspectives that form a connection between nutritional signals and the development of new therapeutic

agents [5,6]. Although little is known about water-soluble vitamins as genetic modulators, there are increasing examples of their effects on gene expression [7–9].

Biotin is a hydrosoluble vitamin that acts as a prosthetic group of carboxylases. Unrelated to its role as carboxylase prosthetic group, biotin regulates gene expression [9,10] and has a wide repertoire of effects on systemic processes such as development [11,12] and immunity [13,14].

The first evidence that biotin affects glucose metabolism was reported by Dakshinamurti et al. [15] in biotin-deficient rats. They observed that glucose tolerance test curves in biotin-deficient rats were significantly higher than those in nondeficient rats, as well as glucose phosphorylation and incorporation into glycogen in the liver [15]. They further demonstrated that these effects were the result of a reduction in the activity of hepatic glucokinase [16], a critical enzyme that regulates glucose uptake in the liver. It has now been well established that the effect of biotin on liver glucokinase activity is not limited to biotin-deficient rats and that such stimulation is observed in rats receiving adequate and pharmacological concentrations of biotin in the diet [16]

[☆] This paper was presented at the international symposium “Vitamins as Regulators of Genetic Expression: Biotin as a Model” NAFTA Satellite Meeting to the XXV National Congress of Biochemistry” held on December 3–4, 2004, in Ixtapa, Zihuatanejo, Mexico. This meeting was sponsored by Sociedad Mexicana de Bioquímica A.C.; Programa de Doctorado en Ciencias Biomedicas, Universidad Nacional Autonoma de Mexico; Laboratorios Roche–Syntex, Mexico; and Instituto de Investigaciones Biomedicas, Universidad Nacional Autonoma de Mexico.

* Tel.: +52 55 56 06 35 58; fax: +52 55 56 06 34 89.

E-mail address: crisfern@biomedicas.unam.mx

as well as in diabetic rats [17]. The effect of biotin on glucokinase is also observed in cultured rat hepatocytes [18] and appears to be mediated through biotin-induced increases of cyclic GMP [18]. Later, with the arrival of molecular biology techniques, Chauhan and Dakshinamurti [19] demonstrated that the stimulatory effect of biotin occurs at the level of transcription. This important discovery set the hallmark on the studies of the role of biotin in gene expression.

Since the finding that biotin increases hepatic glucokinase transcription [19], significant progress has been made in the identification of genes whose expressions are affected by biotin (reviewed in Ref. [9]). The vitamin regulates genes that are critical in the regulation of intermediary metabolism. We [20] found that biotin stimulates the expression of insulin and pancreatic glucokinase, an enzyme that plays an important role in glucose homeostasis regulating insulin secretion in response to changes in blood glucose concentrations. Our studies found that biotin concentrations of 10 to 1000 nM augmented glucokinase activity and mRNA abundance in cultured pancreatic islets isolated from non-biotin-deficient rats [20]. A similar stimulatory effect on pancreatic glucokinase was observed in the insulinoma RIN 1046-38 cell line [21]. We also found that insulin expression and secretion were increased in response to biotin [20]. On the other hand, we found that biotin deficiency decreased pancreatic glucokinase activity and mRNA abundance: Islets isolated from biotin-deficient rats showed about half of the glucokinase activity and mRNA levels observed in control rats. Furthermore, insulin secretion in response to glucose was also impaired in islets isolated from deficient rats [20]. Another essential protein involved in intermediary metabolism, insulin receptor, has also been shown to be regulated by biotin: In the hepatoblastoma cell line HuH7, De la Vega and Stockert [22] found that biotin increased the insulin receptor at the posttranscriptional level; they also found that this effect required the activation of the cGMP signal cascade. Contrary to its stimulatory effects on genes whose action favors hypoglycemia (insulin, insulin receptor, pancreatic and hepatic glucokinase), biotin decreases the expression of hepatic phosphoenolpyruvate carboxykinase [23], a key gluconeogenic enzyme that stimulates glucose production by the liver and thus opposes the glucose catabolic pathways involving hepatic glucokinase.

As can be expected due its role as a prosthetic group of carboxylases, biotin availability modulates the activity of pyruvate carboxylase (PC), a pivotal enzyme in anaplerosis and gluconeogenesis, and acetyl-CoA carboxylases 1 and 2 (ACC 1, ACC 2), two enzymes that regulate fatty acid synthesis and oxidation, respectively. In addition, biotin also regulates the genetic expression of these enzymes at the mRNA level in the case of ACC 1 [24] and at the posttranscriptional level in the case of PC [25].

The findings that biotin regulates the expression of genes that are critical in the regulation of intermediary

metabolism are in agreement with several observations that indicate that biotin supply is involved in glucose and lipid homeostasis.

Biotin deficiency has been linked to impaired oral glucose tolerance tests and decreased utilization of glucose in rats [15,26]. In diabetic patients, it has been found that the biotin status is also altered. Studies by Maebashi et al. [27] in Type 2 diabetic patients found that serum biotin concentration was lower than in control subjects. An inverse correlation between serum biotin and fasting blood glucose concentration has also been observed [27,28]. However, we found that lymphocyte PCC activity, currently considered a more accurate indicator than plasma biotin concentration [29,30], did not differ significantly between mild hyperglycemic Type 2 diabetic patients and nondiabetic subjects [31]. In this work, we also found that pharmacological doses of 15 mg/day of biotin increased lymphocyte PC, ACC and PCC activities. Twenty-eight days of treatment in subjects, either diabetic or nondiabetic, who received biotin increased PCC and ACC activities by approximately 100% and PC activity by approximately 200%.

On the other hand, biotin excess appears to ameliorate the diabetic state. In genetically diabetic KK mice and in OLETF rats, biotin treatment lowered postprandial glucose concentration and improved tolerance to glucose [32,33]. The hypoglycemic effect of the vitamin has also been observed in humans. Reduced hyperglycemia was observed in a group of Type 1 diabetic patients receiving 16 mg/day of biotin for 1 week [28]. This improvement has also been observed in Japanese Type 2 diabetic patients in whom a decrease of about 45% hyperglycemic fasting blood glucose concentrations was observed after 1 month of treatment with oral doses of 9 mg/day of biotin [27]. In hemodialysis patients, pharmacological doses of biotin improved their oral glucose tolerance tests [34]. Studies in our laboratory [35] found that 28 days of treatment with 15 mg of biotin decreased glucose and insulin concentrations.

Several studies have reported a relationship between biotin and lipid metabolism [36–40]. Spontaneous symptoms of biotin deficiency were detected in rats genetically prone to development of elevated blood lipids [36]. In this rat strain, an inverse association was found between plasma lipids and biotin status [37]. A negative correlation between plasma biotin concentrations and blood lipids was also found in humans [38]. Furthermore, pharmacological doses of biotin modify plasma lipid concentrations: A decrease in plasma lipids was observed in humans within 30 min of infusion of 100 mg of biotin [39]. In healthy volunteers, oral biotin supplementation affected plasma lipid concentrations [39]. In patients with atherosclerosis and hyperlipidemia, Dukusova and Krivoruchenko [40] found that the administration of 5 mg/day of biotin for 4 weeks decreased hypercholesterolemia. We [31] found that biotin treatment (15 mg/day) for 28 days decreased hypertriglyceridemia in all subjects (four diabetics and one nondiabetic) whose triacylglycerol concentrations were higher than 25% above

the normal standard of 1.80 mmol/L. These observations support the concept that biotin is able to decrease exacerbated hyperlipidemia.

Little is known about the effects of pharmacological doses of biotin on the expression of lipid metabolism enzymes. In rats, Lewis et al. [41] found that in rats consuming a diet containing 100 mg/kg of biotin during 21 days, the abundance of biotinylated ACC 2 but not that of ACC 1 mass significantly reduced. ACC 2 inhibition has been shown to lead to a decrease in malonyl-CoA levels and the disinhibition of fatty acid oxidation [42]. Therefore, it may be possible that pharmacological doses of biotin increase fatty acid oxidation by decreasing ACC 2 activity.

The notion that a vitamin can be used as a therapeutic agent is not new (e.g., niacin has been used since 1955 as a lipid-lowering drug beyond its role as a vitamin) [43,44]. Furthermore, extensive research on the molecular mechanisms of lipophilic Vitamins A and D has opened new perspectives that connect nutritional signals and the development of new therapeutic agents [5] including promising candidates in diabetes treatment [45,46]. It is only until recently that the molecular action of biotin and its effects on different metabolic functions have attracted attention. The important role of biotin in systemic functions is being reconsidered with the help of new technologies and with novel results regarding the molecular mechanisms of this vitamin, and, as with Vitamins A and D, this might lead to new perspectives in the development of therapeutic agents. The effects of biotin on carbohydrate metabolism and the lack of toxic effects of the vitamin at pharmacological doses suggest that biotin could be used in the development of new therapeutics in the treatment of hyperglycemia and hyperlipidemia, an area that we are actively investigating.

References

- [1] Carlberg C. Lipid soluble vitamins in gene regulation. *Biofactors* 1999;10:91–7.
- [2] Balmer JE, Blomhoff R. Gene expression regulation by retinoic acid. *J Lipid Res* 2002;43:1773–808.
- [3] Chistakos S, Dhawan P, Liu Y, Peng X, Porta A. New insights into the mechanism of vitamin D action. *J Cell Biochem* 2003;88:695–705.
- [4] Combs Jr GF. Biotin. In: *The vitamins. Fundamental aspects in nutrition and health*. 2nd ed. San Diego: Academic Press Inc; 1994. p. 329–45.
- [5] Hinds TS, West WL, Knight EM. Carotenoids and retinoids: a review of research, clinical, and public health applications. *J Clin Pharmacol* 1997;37:551–8.
- [6] Wu-Wong JR, Tian J, Goltzman D. Vitamin D analogs as therapeutic agents: a clinical study update. *Curr Opin Investig Drugs* 2004; 5:320–6.
- [7] Brandsch R. Regulation of gene expression by cofactors derived from B vitamins. *J Nutr Sci Vitaminol* 1994;40:371–99.
- [8] Tully DB, Allgood VE, Cidlowski JA. Modulation of steroid receptor-mediated gene expression by vitamin B6. *FASEB J* 1994;8:343–9.
- [9] Rodríguez-Melendez R, Zemleni J. Regulation of gene expression by biotin. *J Nutr Biochem* 2003;14:680–90.
- [10] Dakshinamurti K, Chauhan J. Biotin-binding proteins. In: Dakshinamurti K, editor. *Vitamin receptors: vitamins as ligands in cell communication*, vol. 1. Cambridge: University Press; 1994. p. 200–49.
- [11] Watanabe T, Endo A. Teratogenic effects of maternal biotin deficiency on mouse embryos examined at midgestation. *Teratology* 1990;42: 295–300.
- [12] Watanabe T. Morphological and biochemical effects of excessive amounts of biotin on embryonic development in mice. *Experientia* 1996; 52:149–54.
- [13] Báez-Saldaña A, Díaz G, Espinoza B, Ortega E. Biotin deficiency induces changes in subpopulations of spleen lymphocytes in mice. *Am J Nutr* 1998;64:431–7.
- [14] Báez-Saldaña A, Ortega E. Biotin deficiency accelerates thymus involution, blocks thymocyte maturation and decreases nose-rump length in mice. *J Nutr* 2004;134:1979–87.
- [15] Dakshinamurti K, Modi VV, Mistry SP. Some aspects of carbohydrate metabolism in biotin-deficient rats. *Proc Soc Exp Biol Med* 1968;127:396–400.
- [16] Dakshinamurti K, Cheah-Tan C. Biotin-mediated synthesis of hepatic glucokinase in the rat. *Arch Biochem Biophys* 1968;127:17–21.
- [17] Dakshinamurti K, Tarrago-Litvak S, Ho Choang H. Biotin and glucose metabolism. *Can J Biochem* 1970;48:493–500.
- [18] Spence JT, Koudelka AP. Effects of biotin upon the intracellular level of cGMP and the activity of glucokinase in cultured rat hepatocytes. *J Biol Chem* 1984;259:6393–6.
- [19] Chauhan J, Dakshinamurti K. Transcriptional regulation of the glucokinase gene by biotin in starved rats. *J Biol Chem* 1991;266: 10035–8.
- [20] Romero-Navarro G, Cabrera-Valladares G, German MS, Matschinsky FM, Wang J, Fernandez-Mejia C. Biotin regulation of pancreatic glucokinase and insulin in primary cultured rat islets and in biotin deficient rats. *Endocrinology* 1999;140:4595–600.
- [21] Borboni P, Magnaterra R, Rabini RA, Staffolanni R, Porzio O, Sesti G, et al. Effect of biotin on glucokinase activity, mRNA expression and insulin release in cultured beta-cells. *Acta Diabetol* 1996;33: 154–8.
- [22] De la Vega L, Stockert R. Regulation of the insulin and asialoglycoprotein receptors via cGMP-dependent protein kinase. *Am J Physiol Cell Physiol* 2000;279:C2037–42.
- [23] Dakshinamurti K, Li W. Transcriptional regulation of liver phosphoenolpyruvate carboxykinase by biotin in diabetic rats. *Mol Cell Biochem* 1994;132:127–32.
- [24] Solorzano-Vargas S, Pacheco-Alvarez D, Leon-Del-Rio A. Holocarboxylase synthetase is an obligate participant in biotin-mediated regulation of its own expression and of biotin-dependent carboxylases mRNA levels in human cells. *Proc Natl Acad Sci U S A* 2002;99: 5325–30.
- [25] Rodríguez-Melendez R, Cano S, Mendez ST, Velazquez A. Biotin regulates the genetic expression of holocarboxylase synthetase and mitochondrial carboxylases in rats. *J Nutr* 2001;131:1909–13.
- [26] Deodhar AD, Mistry SP. Control of glycolysis in biotin deficient rat liver. *Life Sci* 1970;9:581–8.
- [27] Maebashi M, Makino Y, Furukawa Y, Ohinata K, Kimura S, Takao S. Therapeutic evaluation of the effect of biotin on hyperglycemia in patients with non-insulin diabetes mellitus. *J Clin Biochem Nutr* 1993;14:211–8.
- [28] Coggeshall JC, Heggors JP, Robson MC, Baker H. Biotin status and plasma glucose levels in diabetics. *Ann N Y Acad Sci* 1985;447: 389–92.
- [29] Mock DM, Mock NI. Lymphocyte propionyl-CoA carboxylase is an early and sensitive indicator of biotin deficiency in rats, but urinary excretion of 3-hydroxypropionic acid is not. *J Nutr* 2002;132:1945–50.
- [30] Velázquez A, Terán M, Báez A, Gutierrez J, Rodríguez R. Biotin supplementation affects lymphocyte carboxylases and plasma biotin in severe protein-energy malnutrition. *Am J Clin Nutr* 1995;61:385–91.
- [31] Báez-Saldaña A, Zendejas-Ruiz I, Revilla-Monsalve C, Islas-Andrade S, Cárdenas A, Rojas-Ochoa A, et al. Effects of biotin on pyruvate carboxylase, acetyl-CoA carboxylase, propionyl CoA carboxylase, and markers for glucose and lipid homeostasis in type 2 diabetic patients and in nondiabetic subjects. *Am J Clin Nutr* 2004;79:238–43.

- [32] Reddi A, DeAngelis B, Frank O, Lasker N, Baker H. Biotin supplementation improves glucose and insulin tolerances in genetically diabetic KK mice. *Life Sci* 1988;42:1323–30.
- [33] Zhang H, Osada K, Maebashi M, Ito M, Komai M, Furukawa Y. A high biotin diet improves the impaired glucose tolerance of long-term spontaneously hyperglycemic rats with non-insulin-dependent diabetes mellitus. *J Nutr Sci Vitaminol* 1996;42:517–26.
- [34] Koustikos D, Fourtounas C, Kapetanaki A, Agroyannis B, Tzanatos H, Rammos G, et al. Oral glucose test after high-dose i.v. biotin administration in normoglycemic hemodialysis patients. *Ren Fail* 1966;18:131–7.
- [35] Cristina Fernandez-Mejia C, Zendejas-Ruiz I, Revilla Monsalve C, Islas-Andrade S, Báez-Saldaña A, Cárdenas A, et al. Biotin treatment increases insulin sensitivity in type 2 diabetics. American Diabetes Association 63rd Scientific Sessions. *Diabetes* 2003;52(Suppl):A459.
- [36] Marshall MW, Smith BP, Lehman RP. Dietary response of two genetically different lines of inbred rats: lipids in serum and liver. *Proc Soc Exp Biol Med* 1969;131:1271–7.
- [37] Marshall MW, Haubrich M, Washington VA, Chang MW, Young CW, Wheeler MA. Biotin status and lipid metabolism in adult obese hypercholesterolemic inbred rats. *Nutr Metab* 1976;20:41–61.
- [38] Marshall MW, Kliman PG, Washington VA. Effects of biotin on lipids and on other constituents of plasma of healthy men and women. *Artery* 1980;7:330–51.
- [39] Steigerwal H, Bohele H. On the influence of biotin upon the intermediated metabolism. *Internat. Sympos. on drugs affecting lipid metab*, June 2–4; Milan, Italy; 1960. p. 484–6.
- [40] Dukusova OD, Krivoruchenko IV. The effect of biotin on the blood cholesterol levels of atherosclerotic patients in idiopathic hyperlipidemia. *Kardiologia* 1972;12:113.
- [41] Lewis B, Rathman S, McMahon R. Dietary biotin intake modulates the pool of free and protein-bound biotin in rat liver. *J Nutr* 2001;131:2310–5.
- [42] Munday MR, Hemingway CJ. The regulation of acetyl-CoA carboxylase — a potential target for the action of hypolipidemic agents. *Adv Enzyme Regul* 1999;39:205–34.
- [43] Ganji SH, Kamanna VS, Kashyap ML. Niacin and cholesterol: role in cardiovascular disease (review). *J Nutr Biochem* 2003;14:298–305.
- [44] Ito MK. Niacin-based therapy for dyslipidemia: past evidence and future advances. *Am J Manag Care* 2002;8:S315–322.
- [45] Mukherjee R, Davies P, Crombie DL, Bischoff ED, Cesario RM, Jow L, et al. Sensitization of diabetic and obese mice to insulin by retinoic X receptor agonist. *Nature* 1997;386:407–10.
- [46] Liu YL, Sennitt MV, Hislop DC, Crombie DL, Heyman RA, Cawthorne MA. Reginoid X receptor agonists have anti-obesity effects and improve insulin sensitivity in Zucker *fa/fa* rats. *Int J Obes Relat Metab Disord* 2000;24:997–1004.